Excessive dietary lead reduces growth performance and increases lead accumulation in pigs

Hyunjun *Choi*[1,a], Sang Yun *Ji*[2,a], Hyunwoong *Jo*[1], Minho *Song*[3], and Beob Gyun *Kim*[1,*]

*** Corresponding author:** Beob Gyun Kim

Tel: +82-2-2049-6255, **E-mail:** bgkim@konkuk.ac.kr

[1] Department of Animal Science and Technology, Konkuk University, Seoul, Korea

[2] Animal Nutritional Physiology Team, National Institute of Animal Science, Rural Development Administration, Wanju, Korea

[3] Department of Animal Science and Biotechnology, Chungnam National University, Daejeon, Korea

[a] These authors contributed equally to this work.

ORCID

Hyunjun Choi

https://orcid.org/0000-0000-0000-0000

Sang Yun Ji

https://orcid.org/0000-0000-0000-0000

Hyunwoong Jo

https://orcid.org/0000-0000-0000-0000

Minho Song

https://orcid.org/0000-0000-0000-0000

Beob Gyun Kim

https://orcid.org/0000-0000-0000-0000

Title of the manuscript: Excessive dietary lead reduces growth performance and increases lead accumulation in pigs

ABSTRACT

Objective: The objective of this study was to investigate the influence of dietary lead (Pb) supplementation and feeding period on growth performance, organ weight, and Pb accumulation in pigs.

Methods: In a 56-day feeding experiment, a total of 48 barrows with initial body weight 10.4±0.6 kg were allotted to 2 dietary treatments (0 and 200 mg/kg of supplemental Pb) in a completely randomized design with 6 replicates. Body weight and feed intake were recorded to calculate growth performance. At the end of each 14 day-period (on days 14, 28, 42, and 56), an animal was randomly selected from each pen and slaughtered to collect blood samples, hair samples, 5th rib, heart, liver, kidneys, lungs, and longissimus dorsi muscle samples.

Results: Average daily gain and average daily feed intake were reduced ($p<0.01$) by supplemental Pb during the days 0 to 56. Relative kidney weight to body weight was linearly increased with increasing feeding period in pigs fed the Pb-supplemented diet, but not in pigs fed the control diet ($p<0.05$). The Pb concentrations in hair, 5th rib, kidneys, and lungs were linearly increased with longer feeding period in pigs fed the Pb-supplemented diet, but not in pigs fed the control diet ($p<0.01$).

Conclusion: Dietary Pb supplementation caused growth retardation and Pb accumulation in most organs, particularly in hair, bone, and kidneys in a time-dependent manner.

Keywords: Exposure Time; Lead Accumulation; Organ; Swine; Tissue; Toxicity

INTRODUCTION

Contamination of heavy metals in animal feeds is a problem in animal production and health. Among heavy metals, lead (Pb) exposure in domestic animals due to environmental pollution has been often reported [1]. Contamination of Pb in animal feeds is the major route of Pb exposure in domestic animals [2]. Lead, even at a relatively low concentration, can cause various damages to animals such as poisoning and growth retardation. Death can occur if the contamination is high enough [3,4]. In animals fed with a Pb-contaminated diet, Pb is mainly absorbed through the gastrointestinal tract and accumulated in bone, liver, kidneys, and hair [5,6]. In addition, Pb accumulated in organs and tissues of animals could be exposed to humans through the food chain [2].

The Pb concentrations in organs and tissues are known to be increased with increasing exposure time or Pb concentration in animal feeds [5]. In addition, Pb can accumulate more in young animals than in old ones [7,8]. However, very limited information is available on dietary Pb toxicity in young pigs. Therefore, the objective of this study was to investigate the influence of dietary Pb supplementation and feeding period on growth performance, organ weight, and Pb accumulation in young pigs.

MATERIALS AND METHODS

Animal care

The present experiment was reviewed and approved by the Institutional Animal Care and Use Committee of Konkuk University (KU17123). The animal experiment was performed in accordance with the National Institute of Health Guideline for the Care and Use of Laboratory Animals [9].

Animals, diets, and experimental design

A total of 48 weaned barrows ([Landrace×Yorkshire]×Duroc) with initial body weight (BW) of 10.4±0.6 kg were used to investigate the influence of dietary Pb supplementation on pigs. The animals were allotted to 2 dietary treatments (supplemental Pb at 0 and 200 mg/kg as Pb acetate) in a completely randomized design using a spreadsheet program developed by Kim and Lindemann [10]. Experimental diets were prepared as a 2-phase feeding program (days 0 to 21 and days 21 to 56; Table 1). The diets were mainly based on corn and soybean meal and were formulated to meet or exceed the nutrient requirement estimates suggested by the NRC [11]. Four pigs were housed in each pen (2.0×2.2 m^2) that was equipped with a 2-hole feeder and a nipple drinker. Pigs had free access to feed and water throughout the experiment.

Data and sample collection

Individual BW and feed consumption in each pen were recorded every 14 days (on days 0, 14, 28, 42, and 56) to calculate average daily gain (ADG), average daily feed intake (ADFI), and gain to feed ratio (G:F). Individual feed intake of an animal died on day 2 was estimated using the procedure suggested by Lindemann and Kim [12]. On days 14, 28, 42, and 56, an animal randomly selected from each pen was slaughtered to collect blood samples, hair samples, 5th rib, heart, liver, kidneys, lungs, and longissimus dorsi muscle (LM) samples. Blood samples were collected from the jugular vein with EDTA tubes and stored at 4°C. The organs (heart, liver, kidneys, and lungs) were weighed. The samples except blood were stored in a freezer at −20°C. The relative organ weight to BW was calculated to compensate BW effects.

Chemical analysis

Diets were finely ground and analyzed for gross energy using a bomb calorimeter (Parr 1261; Parr Instrument Co., Moline, IL, USA). Dry matter (method 930.15), crude protein (method 990.03), ether extract (method 920.39), neutral detergent fiber (method 2002.04), acid detergent fiber (method 973.18), ash (method 942.05), calcium (method 978.02), and phosphorus (method 946.06) in the diet were analyzed as described in AOAC [13]. Diet samples were digested [14] and analyzed for Pb by inductively coupled plasma optical emission spectrometry (Optima 8300; PerkinElmer, Waltham, MA, USA). Lead in blood samples was measured by inductively coupled plasma spectroscopy (Agilent 7900; Agilent Technology, Santa Clara, CA, USA) using a method described by Nunes et al [15]. The 5th rib, heart, liver, kidneys, lungs, and LM were dried at 105°C using an air-forced drying oven. After drying, samples were finely ground. Before digestion, hair samples were cleaned [16]. The 5th rib was digested as described by Casteel et al [17]. The samples (hair, heart, liver, kidneys, lungs, and LM) were digested and analyzed following the published procedure [18] with minor modification. Briefly, weighed samples (0.2 g) were placed with 2.5 mL of concentrated HNO_3 and 0.5 mL of concentrated HCl in a Pyrex glass tube. The tubes packed by a screw cap were kept in a water bath at 85°C for 3 h. After digestion, the tubes were kept at room temperature to cool down, and then the digested solution was filtered through a syringe filter with 0.20 μm of pore diameter. Each sample was diluted to 50 mL with double-distilled water in a volumetric flask. The digested samples were analyzed for Pb by the inductively coupled plasma method (Agilent 7900; Agilent Technology).

Statistical analysis

Experimental data were analyzed using the MIXED procedure of SAS (SAS Inst. Inc., Cary, NC, USA). In the statistical model, only diet was included as the fixed variable for performance data while both diet and feeding period were used as fixed variables for other measurements including organ weight and Pb concentration in organs and tissues. Least square means of each treatment were calculated. Orthogonal polynomial contrasts were used to test the effects of dietary Pb supplementation, feeding periods, and the interaction between dietary Pb supplementation and feeding period. An experimental unit was a pen for growth performance and a pig for organ weight and Pb concentration in organs and tissues [19]. Statistical significance and tendency were determined at $p<0.05$ and $0.05 \leq p < 0.10$, respectively.

RESULTS

During the experimental period, all pigs consumed experimental diets well and remained healthy except that one pig in the control group died on day 2. Data from the pig died were excluded in the final calculations and statistical analyses.

The ADG, ADFI, or G:F was not affected by supplemental Pb during days 0 to 14 and days 14 to 28 (Table 2). However, pigs fed a diet supplemented with Pb tended to show decreased final BW ($p = 0.091$), ADG ($p = 0.081$), and ADFI ($p = 0.067$) during days 28 to 42 compared with those of the control group. Final BW and ADFI were decreased ($p<0.05$) by supplemental Pb during days 42 to 56. The ADG and ADFI were decreased ($p<0.01$) by supplemental Pb during days 0 to 56. However, dietary Pb supplementation had no effect on G:F during the overall period.

There was no interaction between dietary Pb supplementation and feeding period for the weight of heart, liver, kidneys, or lungs (Table 3). However, relative kidney weight to BW was linearly increased with increasing feeding period in pigs fed the Pb-supplemented diet but not in pigs fed the control diet ($p<0.05$).

Supplemental Pb at 200 mg/kg resulted in increased Pb concentrations ($p<0.001$) in hair, 5th rib, blood, liver, kidneys, lungs, and LM of pigs (Table 4). In pigs fed the Pb-supplemented diet, Pb concentrations in hair, 5th rib, kidneys, and lungs were linearly increased with longer feeding period but not in pigs fed the control diet ($p<0.01$). When Pb weight in organs were calculated, supplemental Pb resulted in greater ($p<0.001$) Pb weight in liver, kidneys, and lungs. In the Pb-supplemented group, the Pb weight in liver, kidneys, and lungs were linearly increased with longer feeding period but not in the control group ($p<0.05$).

DISCUSSION

The present work revealed that growth performance of pigs fed the Pb-supplemented diet was compromised compared with that of pigs fed the control diet, in agreement with results in previous studies [20,21]. However, some researchers failed to find the negative effects of dietary Pb on growth performance of pigs [22,23]. The lack of responses in performance was likely due to the low dose of Pb, experimental period, or both. While dietary Pb concentration was 200 mg/kg in the present work, in the study by Zacharias et al [23], Pb concentration in the experimental diet was 1.45 mg/kg and feed intake was restricted. The feeding period might be another factor that influences performance responses to dietary Pb. Although the Pb concentration was 250 mg/kg in the study of Reddy et al [22], the feeding period was only 28 days.

The increased kidney weight by dietary Pb supplementation in the present work agrees with results of previous studies using mice and rats [24,25]. Urinary excretion of absorbed Pb is one of major Pb excretion routes, which indicates that kidneys are target organs for Pb toxicity [25]. In rats, supplemental Pb can reduce the concentration of glutathione and antioxidant enzymes [24] but increase cell proliferation in the proximal tubular epithelium of the kidney [26]. These effects of Pb toxicity on the kidney may be a major reason for the increased kidney weight in the Pb-supplemented group observed in the present work.

While the absorption rate of Pb is less than other heavy metals, Pb is relatively slowly excreted from animals. Therefore, Pb is accumulated well in most tissues once absorbed into animal body [8]. Previous studies have reported that Pb is highly accumulated in bone, kidneys, liver, and hair [20,21], but not in muscle [23,27], which agrees with the present study.

In Pb toxicity experiments, the Pb dose and feeding period are important factors. In the present work, only 2 concentrations (0 and 200 mg/kg) of Pb were used, making it impossible to assess dose-dependent polynomial effects. However, 4 feeding periods (14, 28, 42, and 56 days) were employed and time-dependent effects of dietary Pb were observed. In agreement, previous studies reported that Pb concentrations in organs were increased as the feeding period was increased to 84 days in rats [28] and pigs [27]. In rats, Pb concentrations in tissues were increased with increasing Pb intake and the total amount of Pb in tissue did not affect absorption of Pb [7], which indicates that animals perhaps do not regulate Pb absorption or excretion. Although no data are available on Pb absorption or excretion in pigs, the increased Pb concentrations in pig organs, particularly in bone, hair, and kidneys, by extended feeding of Pb in the present work were likely due to the inability of pigs for excreting

absorbed Pb. Lead elimination rate in bone is less than that in other tissues in rats [7] and pigs [27], which explains the greatest Pb accumulation in bone and the linear response of Pb concentration with increasing exposure time in the present work.

The quadratic increase of Pb concentration in organs with increasing feeding period may be associated with the age of animals and a dietary milk product. Sharma et al [27] have reported that Pb is more highly accumulated in younger pigs. Similarly, as the age of rats increased, the absorption of Pb decreased [7,29]. In the present work, Pb was more highly accumulated in pigs likely due to the young age during the first few weeks. Dietary lactose is also a factor that influences the absorption of Pb. In rats, dietary Pb was more highly accumulated in organs when a lactose-added diet was provided compared with a glucose-added diet [29]. Bell and Spickett [30] have also reported that dietary Pb is more efficiently accumulated in rats fed a dried whole milk diet which contains an appreciable amount of lactose than those fed a lactose-hydrolyzed milk diet. In the present work, dried whey containing lactose was included at 10% in the experimental diets during the first 3 weeks. Dietary lactose may be a potential reason for the relatively high Pb concentrations in the day 14 samples of bone, liver, and kidneys of pigs fed a Pb-supplemented diet.

CONCLUSION

Dietary Pb supplementation can cause growth retardation and increase kidney weight with an increasing feeding period. In addition, when duration of dietary Pb exposure was increased, Pb concentration was increased in organs and tissues, although its accumulation rates varied depending on organs and tissues of pigs. Lead was accumulated particularly in hair, bone, and kidneys in a time-dependent manner.

CONFLICT OF INTEST

The authors declare that there is no conflict of interests.

AUTHORS' CONTRIBUTION

Conceptualization: Choi H, Kim BG.

Data curation: Choi H, Yun S.

Formal analysis: Jo H.

Methodology: Choi H, Yun S.

Software: Jo H, Song M.

Validation: Song M, Kim BG.

Investigation: Choi H, Yun S.

Writing - original draft: Choi H, Kim BG.

Writing - review & editing: Choi H, Yun S, Jo H, Song M, Kim BG.

FUNDING

The authors are grateful for the support by Rural Development Administration (Republic of Korea; PJ010932).

ACKNOWLEGEMENTS

Not applicable.

SUPPLEMENTARY MATERIAL

Not applicable.

DATA AVAILABILITY

Upon reasonable request, the datasets of this study can be available from the corresponding author.

ETHICS APPROVAL

The present experiment was reviewed and approved by the Institutional Animal Care and Use Committee of Konkuk University (KU17123). The animal experiment was performed in accordance with the National Institute of Health Guideline for the Care and Use of Laboratory Animals

DECLARATION OF GENERATIVE AI

No AI tools were used in this article.

REFERENCES

1. Lockitch G. Perspectives on lead toxicity. Clin Biochem 1993;26:371-81. https://doi.org/10.1016/0009-

9120(93)90113-K

2. Brams E, Anthony W. Cadmium and lead through an agricultural food chain. Sci Total Environ 1983;28:295-306. https://doi.org/10.1016/S0048-9697(83)80027-8
3. Doyle JJ, Spaulding JE. Toxic and essential trace elements in meat – a review. J Anim Sci 1978;47:398-419. https://doi.org/10.2527/jas1978.472398x
4. Gurer H, Ercal N. Can antioxidants be beneficial in the treatment of lead poisoning? Free Radic Biol Med 2000;29:927-45. https://doi.org/10.1016/S0891-5849(00)00413-5
5. Mertz W. Trace elements in human and animal nutrition. 5th ed. Academic Press; 1986.
6. Tariq H, Sharma A, Sarkar S, et al. Perspectives for rare earth elements as feed additive in livestock – a review. Asian-Australas J Anim Sci 2020;33:373-81. https://doi.org/10.5713/ajas.19.0242
7. Conrad ME, Barton JC. Factors affecting the absorption and excretion of lead in the rat. Gastroenterology 1978;74:731-40. https://doi.org/10.1016/0016-5085(78)90253-6
8. Humphreys DJ. Effects of exposure to excessive quantities of lead on animals. Br Vet J 1991;147:18-30. https://doi.org/10.1016/0007-1935(91)90063-S
9. National Institutes of Health, National Research Council. Guide for the care and use of laboratory animals. 8th ed. National Academies Press; 2010.
10. Kim BG, Lindemann MD. A new spreadsheet method for the experimental animal allotment. J Anim Sci 2007;85(Suppl. 2):112.
11. Committee on Nutrient Requirements of Swine, National Research Council. Nutrient requirements of swine. 11th ed. National Academies Press; 2012.
12. Lindemann MD, Kim BG. A model to estimate individual feed intake of swine in group feeding. J Anim Sci 2007;85:972-5. https://doi.org/10.2527/jas.2006-412
13. Association of Official Analytical Chemists (AOAC) International. Official methods of analysis of AOAC International. 18th ed. AOAC International; 2005.
14. Wu S, Feng X, Wittmeier A. Microwave digestion of plant and grain reference materials in nitric acid or a mixture of nitric acid or a mixture of nitric acid and hydrogen peroxide for the determination of multi-elements by inductively coupled plasma mass spectrometry. J Anal At Spectrom 1997;12:797-806. https://doi.org/10.1039/A607217H
15. Nunes JA, Batista BL, Rodrigues JL, et al. A simple method based on ICP-MS for estimation of background

levels of arsenic, cadmium, copper, manganese, nickel, lead, and selenium in blood of the Brazilian population. J Toxicol Environ Health Part A 2010;73:878-87. https://doi.org/10.1080/15287391003744807

16. Chaturvedi R, Banerjee S, Chattopadhyay P, et al. High iron accumulation in hair and nail of people living in iron affected areas of Assam, India. Ecotoxicol Environ Saf 2014;110:216-20. https://doi.org/10.1016/j.ecoenv.2014.08.028
17. Casteel SW, Weis CP, Henningsen GM, Brattin WJ. Estimation of relative bioavailability of lead in soil and soil-like materials using young swine. Environ Health Perspect 2006;114:1162-71. https://doi.org/10.1289/ehp.8852
18. Ashoka S, Peake BM, Bremner G, Hageman KJ, Reid MR. Comparison of digestion methods for ICP-MS determination of trace elements in fish tissues. Analytica Chimica Acta 2009;653:191-9. https://doi.org/10.1016/j.aca.2009.09.025
19. Seo S, Jeon S, Ha JK. Guidelines for experimental design and statistical analyses in animal studies submitted for publication in the Asian-Australasian Journal of Animal Sciences. Asian-Australas J Anim Sci 2018;31:1381-6. https://doi.org/10.5713/ajas.18.0468
20. Hsu FS, Krook L, Pond WG, Duncan JR. Interactions of dietary calcium with toxic levels of lead and zinc in pigs. J Nutr 1975;105:112-8. https://doi.org/10.1093/jn/105.1.112
21. Phillips C, Gyori Z, Kovacs B. The effect of adding cadmium and lead alone or in combination to the diet of pigs on their growth, carcase composition and reproduction. J Sci Food Agric 2003;83:1357-65. https://doi.org/10.1002/jsfa.1548
22. Reddy KE, Park KR, Lee SD, et al. Effects of graded concentrations of supplemental lead on lead concentrations in tissues of pigs and prediction equations for estimating dietary lead intake. PeerJ 2017;5:e3936. https://doi.org/10.7717/peerj.3936
23. Zacharias B, Lantzsch HJ, Drochner W. Influence of microbial phytase and dietary calcium on the accumulation of lead in different organs of pigs. Biol Trace Elem Res 1999;70:243-9. https://doi.org/10.1007/BF02783833
24. Abdel-Moneim AE, Dkhil MA, Al-Quraishy S. The potential role of flaxseed oil on lead acetate-induced kidney injure in adult male Albino rats. Afr J Biotechnol 2011;10:1436-51. https://doi.org/10.5897/AJB10.1310
25. Mohammadi S, Zamani E, Mohadeth Z, et al. Effects of different doses of simvastatinon lead-induced kidney

damage in Balb/c male mice. Pharm Sci 2015;20:157-62.

26. Choie DD, Richter GW. Cell proliferation in rat kidney induced by lead acetate and effects of uninephrectomy on the proliferation. Am J Pathol 1972;66:265-75.
27. Sharma RP, Street JC, Shupe JL. Translocation of lead and cadmium from feed to edible tissues of swine. J Food Saf 1982;4:151-63. https://doi.org/10.1111/j.1745-4565.1982.tb00439.x
28. Areola OO, Jadhav AL, Williams-Johnson M. Relationship between lead accumulation in blood and soft tissues of rats subchronically exposed to low levels of lead. Toxic Subst Mech 1999;18:149-61. https://doi.org/10.1080/107691899229115
29. Bushnell PJ, DeLuca HF. The effects of lactose on the absorption and retention of dietary lead. J Nutr 1983;113:365-78. https://doi.org/10.1093/jn/113.2.365
30. Bell RR, Spickett JT. The influence of milk in the diet on the toxicity of orally ingested lead in rats. Food Cosmet Toxicol 1981;19:429-36. https://doi.org/10.1016/0015-6264(81)90446-6

292 **Table 1.** Ingredient and analyzed chemical compositions of control diets (as-fed basis)

Item	Days 0 to 21	Days 21 to 56
Ingredient (%)		
Ground corn	58.26	62.28
Soybean meal (48% crude protein)	24.00	33.00
Dried whey	10.00	-
Fish meal	3.00	-
Soybean oil	2.00	2.00
$_{L}$-Lys·HCl (78.8%)	0.38	-
$_{DL}$-Met (99%)	0.08	-
$_{L}$-Thr (99%)	0.12	-
Dicalcium phosphate	0.48	1.00
Ground limestone	0.88	0.82
Mineral premix[1)]	0.25	0.25
Vitamin premix[2)]	0.25	0.25
Salt	0.30	0.40
Analyzed composition (%)		
Dry matter	90.20	88.80
Gross energy (kcal/kg)	4,025	4,016
Crude protein	19.70	19.10
Ether extract	6.49	6.09
Ash	5.17	4.74
Calcium	0.52	0.52
Phosphorus	0.49	0.34
Neutral detergent fiber	7.51	8.56
Acid detergent fiber	2.39	2.73
Lead[3)] (Pb, mg/kg)	Not detected	Not detected

293 [1)] Provided the following quantities per kg of complete diet: vitamin A, 12,500 IU; vitamin D_3, 1,000 IU; vitamin
294 E, 125 IU; vitamin K_3, 6.3 mg; thiamin, 6.3 mg; riboflavin, 25.0 mg; pyridoxine, 12.5 mg; vitamin B_{12}, 0.1 mg;
295 pantothenic acid, 100 mg; folic acid, 7.5 mg; niacin, 225 mg; and biotin, 0.5 mg.
296 [2)] Provided the following quantities per kg of complete diet: Cu, 87.5 mg as copper sulfate; Fe, 125 mg as iron
297 sulfate; I, 1.0 mg as potassium iodate; Mn, 75 mg as manganese sulfate; Se, 0.25 mg as sodium selenite; and Zn,
298 60 mg as zinc oxide.
299 [3)] The analyzed Pb concentrations in the Pb-supplemented diets were 138.6 and 238.5 mg/kg, respectively, during
300 days 0 to 21 and days 21 to 56.

Table 2. Influence of dietary lead (Pb) supplementation on growth performance of pigs[1)]

Item	Supplemental Pb, mg/kg		SEM	p-value
	0	200		
Days 0 to 14				
Initial body weight, kg	10.28	10.49	0.12	0.226
Final body weight, kg	16.60	16.21	0.42	0.526
Average daily gain, g/d	451	409	27	0.284
Average daily feed intake, g/d	736	683	33	0.289
Gain to feed ratio	0.61	0.60	0.02	0.702
Days 14 to 28				
Initial body weight, kg	16.67	16.30	0.41	0.539
Final body weight, kg	24.43	23.50	0.73	0.390
Average daily gain, g/d	554	514	32	0.391
Average daily feed intake, g/d	1,122	1,046	33	0.139
Gain to feed ratio	0.49	0.49	0.02	0.907
Days 28 to 42				
Initial body weight, kg	24.88	23.60	0.81	0.289
Final body weight, kg	35.08	31.22	1.46	0.091
Average daily gain, g/d	729	545	67	0.081
Average daily feed intake, g/d	1,510	1,298	73	0.067
Gain to feed ratio	0.48	0.41	0.04	0.199
Days 42 to 56				
Initial body weight, kg	34.92	30.58	1.34	0.046
Final body weight, kg	44.83	39.83	1.07	0.008
Average daily gain, g/d	708	661	58	0.572
Average daily feed intake, g/d	2,144	1,626	136	0.022
Gain to feed ratio	0.34	0.43	0.06	0.325
Days 0 to 56				
Average daily gain, g/d	612	518	18	0.004
Average daily feed intake, g/d	1,373	1,137	50	0.007
Gain to feed ratio	0.45	0.46	0.01	0.703

[1)] Each least squares mean represents 6 replicated pens; a pig selected from each pen was slaughtered at the end of each 14-day period.

SEM, standard error of the means.

306 **Table 3.** Influence of dietary lead (Pb) supplementation and feeding period on organ weight of pigs (wet basis)

	Pb, mg/kg:	0				200					p-value[1)]				
Item	Period, day:	14	28	42	56	14	28	42	56	SEM	Pb	L	Q	Pb×L	Pb×Q
No. of observations		6	5	6	6	6	6	6	6						
Organ weight, g															
Heart		88	129	170	210	87	127	145	189	9	0.048	<0.001	0.952	0.133	0.816
Liver		479	627	887	1,033	422	580	744	923	40	0.002	<0.001	0.867	0.295	0.836
Kidneys		81	116	173	219	81	134	180	254	10	0.035	<0.001	0.247	0.127	0.691
Lungs		193	259	353	448	196	303	344	399	34	0.904	<0.001	0.805	0.323	0.395
Organ weight relative to body weight[2)], %															
Heart		0.54	0.56	0.49	0.47	0.54	0.55	0.45	0.47	0.02	0.583	<0.001	0.661	0.800	0.317
Liver		2.92	2.72	2.52	2.30	2.65	2.52	2.36	2.31	0.10	0.028	<0.001	0.800	0.142	0.715
Kidneys		0.49	0.50	0.49	0.49	0.51	0.58	0.57	0.64	0.03	<0.001	0.074	0.880	0.037	0.856
Lungs		1.18	1.14	1.01	1.00	1.23	1.33	1.08	1.00	0.10	0.246	0.010	0.617	0.664	0.470

307 [1)] Pb, dietary Pb supplementation; L, linear effect of feeding period; Q, quadratic effect of feeding period; Pb×L, interaction between dietary Pb supplementation and linear
308 effect of feeding period; Pb×Q, interaction between dietary Pb supplementation and quadratic effect of feeding period.
309 [2)] Relative organ weights to body weight (%) = organ weight (kg)/body weight of pig (kg)×100.
310 SEM, standard error of the means.
311

312 **Table 4**. Influence of dietary lead (Pb) supplementation and feeding period on Pb concentration of pig organs (wet basis except for hair)

Item	Pb, mg/kg:	0				200				SEM	p-value[1)]				
	Period, day:	14	28	42	56	14	28	42	56		Pb	L	Q	Pb×L	Pb×Q
No. of observations		6	5	6	6	6	6	6	6						
Hair, mg/kg		1.42	0.92	1.47	1.36	4.35	9.15	17.63	26.23	2.06	<0.001	<0.001	0.460	<0.001	0.547
5th rib, mg/kg		3.58	1.64	1.68	1.90	44.58	42.68	100.58	149.03	9.35	<0.001	<0.001	0.046	<0.001	0.066
Blood, μg/dL		0.013	0.007	0.003	0.003	0.378	0.582	0.454	0.521	0.031	<0.001	0.178	0.059	0.089	0.042
Pb concentration in fresh tissue[2)], mg/kg															
Heart		0.60	0.70	0.92	0.59	0.81	0.83	0.56	1.19	0.17	0.206	0.286	0.711	0.512	0.027
Liver		0.27	0.49	0.65	1.06	10.49	6.99	8.64	12.14	0.74	<0.001	0.047	0.001	0.368	0.002
Kidneys		0.50	0.23	0.17	0.20	7.95	7.03	8.89	10.09	0.44	<0.001	0.008	0.049	0.001	0.132
Lungs		0.21	0.10	0.10	0.07	0.27	0.31	0.37	0.36	0.04	<0.001	0.780	0.857	0.002	0.245
LM		0.14	0.20	0.13	0.24	0.30	0.26	0.27	0.25	0.03	<0.001	0.663	0.437	0.100	0.715
Pb weight in fresh organ, mg															
Heart		0.05	0.09	0.16	0.12	0.07	0.11	0.09	0.23	0.03	0.433	<0.001	0.675	0.305	0.026
Liver		0.13	0.31	0.57	1.11	4.46	4.04	6.60	11.27	0.61	<0.001	<0.001	0.002	<0.001	0.007
Kidneys		0.04	0.03	0.03	0.04	0.64	0.94	1.65	2.59	0.13	<0.001	<0.001	0.065	<0.001	0.087
Lungs		0.04	0.03	0.04	0.03	0.05	0.09	0.15	0.15	0.02	<0.001	0.022	0.559	0.012	0.444

313 [1)] Pb, dietary Pb supplementation; L, linear effect of feeding period; Q, quadratic effect of feeding period; Pb×L, interaction between dietary Pb supplementation and linear
314 effect of feeding period; Pb×Q, interaction between dietary Pb supplementation and quadratic effect of feeding period.
315 [2)] The values were calculated based on the Pb concentrations in dried organ and moisture concentration in fresh organ.
316 LM, longissimus dorsi muscle; SEM, standard error of the means.
317